

Review

Epithelial Impermeability to Water: A Second Look

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Key Words

Loop of Henle • Renal Collecting Ducts • Water Permeability • Countercurrent Mechanisms

Abstract

Impermeability of the ascending limb of the Henle loop for water is traditionally regarded as essential for countercurrent multiplication in the kidney. Similar claims have been made about permeability properties of the collecting duct and some other epithelia. It is not clear, however, how a structure based on phospholipid bilayers can be water-impermeant if phospholipid bilayers themselves have measurable permeability. The presence of two membranes separated by the cytoplasm may only account for a several-fold reduction in permeability compared to a single bilayer. By analyzing published data, we conclude that these tubules do have a finite water permeability, especially the collecting duct. Although the results on isolated ascending limbs vary among authors, osmotic shock experiments clearly indicate that both the collecting duct and the ascending Henle loop are sufficiently water-permeable to observe volume regulation effects. We conclude that these epithelia by themselves do not display unusual resistance to water flow; it can be estimated that 20-50% of the fluid entering the tubules can be reabsorbed into a strongly hypertonic medulla. It is possible, however, that unstirred layers in the intact kidney may contribute to the apparent low permeability of the tubules.

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Introduction

The belief that the ascending loop of Henle and unstimulated collecting duct are impermeable to water is widely shared by physiologists; it is stated as a matter of course in most textbooks [1-3] and in scientific literature [4-6]. The biological significance of water-impermeant epithelia is that while ions are being removed from the filtrate by various transporters, water is unable to follow ions by osmosis, and the fluid becomes hypotonic. A similar process is responsible for the production of hypotonic saliva [7].

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But even pure lipid membranes are permeable to water to some extent, with osmotic permeabilities P_f in the range of 10 – 160 $\mu\text{m/s}$ [8, 9] (see [10] for the explanation of permeability); how can it be that some epithelia are not? This question has drawn interest from researchers, who pointed out the potential importance of membrane asymmetry, integral proteins, and mucins [11-13]. To that, we would like to add a few additional considerations.

1. Single membrane vs. epithelium.

When comparing the permeability of a phospholipid bilayer with that of a simple epithelium, we must account for the presence of two membranes in the latter (apical and basolateral) separated by cytoplasm. For simplicity, we can assume that both membranes are identical and separated by the distance $h = 7 \mu\text{m}$ [14]; the diffusion coefficient of water D_w in the cytoplasm has been estimated at 400 $\mu\text{m}^2/\text{s}$ [15-17]. For a single membrane, the steady state water flux is expressed as

$$\Phi_m = P_f(C_o - C_i)$$

In the case of a three-layer epithelium, the expression changes to

$$\Phi_e = \frac{P_f D_w}{P_f h + 2D_w}(C_o - C_i)$$

which can be derived by equating water fluxes through each of the three compartments or by using the rule for calculating the combined permeability of membranes in series [18]:

$$\frac{1}{P_{total}} = \sum_i \frac{1}{P_i}$$

Therefore, the additional barriers present in the epithelium slow down water transport by the factor of

$$\frac{\Phi_m}{\Phi_e} = 2 + h \frac{P_f}{D_w}$$

For the values of P_f listed above, this ratio ranges from 2.2 to 4.8. Although this may partly explain the difference between a single membrane and an epithelium (for example, higher values in vesicles isolated from the medullary thick ascending limb (MTAL) than in intact MTAL [18]), most data on kidney permeability have been obtained on isolated tubules.

2. What values of permeabilities would qualify a membrane as water-impermeant?

The rate of water permeation through a tube with length L and radius R and subjected to an osmolarity gradient $\Delta\Pi$ is

$$\Phi_{out} = P_f \cdot 2\pi RL \cdot v_w \cdot \Delta\Pi$$

where v_w is the molar volume of water equal to 18 cm^3/mol . The typical value of P_f for the medullary collecting duct (MCD) in the absence of vasopressin is 20 $\mu\text{m/s}$ [18-20]; some authors have obtained slightly higher [21] or slightly lower [22, 23] values. The permeability of 20 $\mu\text{m/s}$ is regarded as low: " P_f greater than 0.01 cm/sec (at 25–37°C) is considered to be high and suggests the involvement of molecular water channels, whereas P_f less than 0.005

cm/sec is consistent with water diffusion through the lipid portion of a membrane" [24]. However, assuming $R = 12 \mu\text{m}$, $L = 10^4 \mu\text{m}$ [25], and $\Delta\Pi = 0.5 \text{ mol/L}$ during diuresis [26], we find the rate of water removal by a single collecting duct on the order of 6 nl/min, which is comparable to the estimated flow rate $\Phi_{\text{in}} = 14 \text{ nl/min}$ [25]. In other words, almost half of the fluid entering the MCD and destined for elimination is expected to be reabsorbed, even in the "water-impermeable" state of the duct.

The permeability of MTAL has also been a focus of much research. Rocha and Kokko [27] found its permeability close to zero; however, the standard error of their measurements was 4-6 $\mu\text{m/s}$. The results of Sasaki and Imai [28] were similar. The accuracy of those estimates may have been compromised by using fast perfusion rates [29]. Other measurements have produced a wide range of values from 0.07 $\mu\text{m/s}$ [30] to 5 $\mu\text{m/s}$ [23] or between 6 $\mu\text{m/s}$ and 23 $\mu\text{m/s}$ (cited in [29]).

Using the conservative estimate $P_f = 1 \mu\text{m/s}$ and the parameters $R = 15 \mu\text{m}$, $L = 2 \cdot 10^4 \mu\text{m}$, and $\Delta\Pi = 1 \text{ mol/L}$, we find that the leak through the walls would amount to 2 nl/min, or about a quarter of the total flow of 8 nl/min [25]. Here, once again, epithelial impermeability does not directly follow from the data.

3. Cell volume regulation.

A separate body of work has focused on cell volume regulation - restoration of cell water content following either osmotic swelling (regulatory volume decrease, or RVD) or osmotic shrinkage (regulatory volume increase, RVI). The RVI and RVD are secondary responses to osmolarity changes caused by the activation of membrane channels for ions or organic osmolytes [31]; but to initiate these responses, the membrane must be permeable to water in the first place.

The numerous reports of volume regulatory responses in the collecting duct [19, 32-35] and in the ascending loop [36 - 40] provide compelling evidence that these cells are sufficiently permeable to water, as are all other mammalian cells. A single publication claiming the lack of swelling of cheek epithelial cells in hypotonic solutions [41] may have resulted from unnoticed rapid RVD that develops and subsides within a minute (according to our unpublished observations).

Conclusion

The presented brief review suggests that "water-impermeable" kidney epithelia do not possess watertight properties significantly beyond those expected from phospholipid bilayers. Their permeabilities are indeed two orders of magnitude less than the permeability of the thin descending loop [42, 43], which can be due to the lack of water channels, but are similar to those of many other cell types [8, 18, 44-48]. When comparing P_f values for epithelia and isolated lipid layers, the presence of an additional membrane and the cytoplasm should be taken into account; additionally, the effects of unstirred layers can be significant even in perfused tubule preparations [18, 43] and are expected to be particularly prominent in the interstitium of the kidney [49]. Indeed, a several-fold difference between *in vivo* and *in vitro* permeabilities of the salivary duct has been reported [50]. Conceivably, slow convection and dissipation of osmolality gradients can be a factor in water retention.

The main question, however, is whether the diluting function of the nephron and the effective diuresis require that MTAL and unstimulated MCD be strictly water-impermeable. It seems that the existing mathematical models assume zero permeabilities of both the thin and thick ascending loops [51-55], and the possibility of deviations from zero have not been considered. That would be an interesting question to investigate.

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Disclosure Statement

The authors have no conflicts of interest.

References

- Hall JE (2016) Guyton and Hall textbook of medical physiology, 13th ed. Elsevier, Philadelphia.
- Widmaier EP, Raff H, Strang KT (2019) Vander's human physiology, 15th ed. McGraw Hill, New York.
- Silverthorn DU (2019) Human physiology: an integrated approach, 8th ed. Pearson, San Francisco.
- Staruschenko A (2012) Regulation of transport in the connecting tubule and cortical collecting duct. *Compr Physiol* 2:1541.
- Palm F, Carlsson PO (2005) Thick ascending tubular cells in the loop of Henle: regulation of electrolyte homeostasis. *Int J Biochem Cell Biol* 37:1554-1559.
- Randall Thomas S (2009) Kidney modeling and systems physiology. *WIREs Syst Biol Med* 1:172-190.
- Catalán MA, Nakamoto T, Melvin JE (2009) The salivary gland fluid secretion mechanism. *J Med Investig* 56(Suppl):192-196.
- Fettiplace R, Haydon DA (1980) Water permeability of lipid membranes. *Physiol Rev* 60:510-550.
- Mathai JC, Tristram-Nagle S, Nagle JF, Zeidel ML (2008) Structural determinants of water permeability through the lipid membrane. *J Gen Physiol* 131:69-76.
- Manning GS, Kay AR (2023) The physical basis of osmosis. *J Gen Physiol* 155:e202313332.
- Zeidel M (1996) Low permeabilities of apical membranes of barrier epithelia: what makes watertight membranes watertight? *Am J Physiol Renal Physiol* 271:F243-F245.
- Verkman AS (1989) Mechanisms and regulation of water permeability in renal epithelia. *Am J Physiol Cell Physiol* 257:C837-C850.
- Hu P, Meyers S, Liang FX, Deng FM, Kachar B, Zeidel M, Sun TT (2002) Role of membrane proteins in barrier function: uropodin deficit compromises the urothelial permeability barrier. *Am J Physiol* 283:F1200-1207.
- Kumaran GK, Hanukoglu I (2020) Identification and classification of epithelial cells in nephron segments by actin cytoskeleton patterns. *FEBS J* 287:1176-1194.
- Tanner JE (1983) Intracellular diffusion of water. *Arch Biochem Biophys* 224:416-428.
- Rorschach HE, Lin C, Hazlewood CF (1991) Diffusion of water in biological tissues. *Scanning Microsc* 1991:1.
- Mastro AM, Keith AD (1984) Diffusion in the aqueous compartment. *J Cell Biol* 99:180s-187s.
- Rivers R, Blanchard A, Eladari D, Levie F, Paillard M, Pódevin RA, Zeidel ML (1998) Water and solute permeabilities of medullary thick ascending limb apical and basolateral membranes. *Am J Physiol Renal Physiol* 274:F453-F462.
- Strange K, Spring KR (1987) Cell membrane water permeability of rabbit cortical collecting duct. *J Membr Biol* 96:27-43.
- Hebert SC, Schafer JA, Andreoli TE (1981) The effects of antidiuretic hormone (ADH) on solute and water transport in the mammalian nephron. *J Membr Biol* 58:1-9.
- Nielsen S, Chou CL, Marples D, Christensen EI, Kishore BK, Knepper MA (1995) Vasopressin increases water permeability of kidney collecting duct by inducing translocation of aquaporin-CD water channels to plasma membrane. *Proc Natl Acad Sci USA* 92:1013-1017.
- Schafer JA, Andreoli TE (1972) Cellular constraints to diffusion: The effect of antidiuretic hormone on water flows in isolated mammalian collecting tubules. *J Clin Invest* 51:1264-1278.

- 23 Morgan T, Berliner RW (1968) Permeability of the loop of Henle, vasa recta, and collecting duct to water, urea, and sodium. *Am J Physiol (Legacy Content)* 215:108-115.
- 24 Verkman AS (2000) Water permeability measurement in living cells and complex tissues. *J Membr Biol* 173:73-87.
- 25 Gilmer GG, Deshpande VG, Chou CL, Knepper M (2018) Flow resistance along the rat renal tubule. *Am J Physiol Renal Physiol* 315:F1398-F1405.
- 26 Sadowski J, Dobrowolski L (2003) The renal medullary interstitium: focus on osmotic hypertonicity. *Clin Exp Pharmacol Physiol* 30:119-126.
- 27 Rocha AS, Kokko JP (1973) Sodium chloride and water transport in the medullary thick ascending limb of Henle. Evidence for active chloride transport. *J Clin Invest* 52:612-623.
- 28 Sasaki S, Imai M (1980) Effects of vasopressin on water and NaCl transport across the *in vitro* perfused medullary thick ascending limb of Henle's loop of mouse, rat, and rabbit kidneys. *Pflügers Arch* 383:215-21.
- 29 Burg MB (1982) Thick ascending limb of Henle's loop. *Kidney Int* 22:454-464.
- 30 Burg MB, Green NO (1973) Function of the thick ascending limb of Henle's loop. *Am J Physiol (Legacy Content)* 224:659-668.
- 31 Lang F (2007) Mechanisms and significance of cell volume regulation. *J Am Coll Nutr* 26:613S-623S.
- 32 Fu WJ, Kuwahara M, Gragoe Jr EJ, Marumo F (1993) Mechanisms of regulatory volume increase in collecting duct cells. *Jpn J Physiol* 43:745-757.
- 33 Sun A, Hebert SC (1989) Rapid hypertonic cell volume regulation in the perfused inner medullary collecting duct. *Kidney Int* 36:831-842.
- 34 Ford P, Rivarola V, Chara O, Blot-Chabaud M, Cluzeaud F, Farman N, Parisi M, Capurro C (2005) Volume regulation in cortical collecting duct cells: role of AQP2. *Biol Cell* 97:687-697.
- 35 Solenov EI (2008) Cell volume and sodium content in rat kidney collecting duct principal cells during hypotonic shock. *J Biophys* 2008:420963.
- 36 Roger F, Martin PY, Rousselot M, Favre H, Féraïlle E (1999) Cell shrinkage triggers the activation of mitogen-activated protein kinases by hypertonicity in the rat kidney medullary thick ascending limb of the Henle's loop: requirement of p38 kinase for the regulatory volume increase response. *J Biol Chem* 274:34103-34110.
- 37 Onuchic LF, Arenstein IR, Lopes AG (1992) Cell volume regulation in rat thin ascending limb of Henle's loop. *Am J Physiol Renal Physiol* 263:F353-F362.
- 38 Montrose-Rafizadeh CH, Guggino WB (1991) Role of intracellular calcium in volume regulation by rabbit medullary thick ascending limb cells. *Am J Physiol Renal Physiol* 260:F402-F409.
- 39 Grunewald RW, Fahr M, Fiedler GM, Jehle PM, Müller GA (2001) Volume regulation of thick ascending limb of Henle cells: significance of organic osmolytes. *Exp Nephrol* 9:81-89.
- 40 Hebert SC, Sun A (1988) Hypotonic cell volume regulation in mouse medullary thick ascending limb: effects of ADH. *Am J Physiol Renal Physiol* 255:F962-F969.
- 41 Lee EJ, Patten GS, Burnard SL, McMurchie EJ (1994) Osmotic and other properties of isolated human cheek epithelial cells. *Am J Physiol Cell Physiol* 267:C75-C83.
- 42 Chou CL, Knepper MA (1992) *In vitro* perfusion of chinchilla thin limb segments: segmentation and osmotic water permeability. *Am J Physiol Renal Physiol* 263:F417-F426.
- 43 Berry CA (1983) Water permeability and pathways in the proximal tubule. *Am J Physiol Renal Physiol* 245:F279-F294.
- 44 Capurro C, Escobar E, Ibarra C, Porta M, Parisi M (1989) Water permeability in different epithelial barriers. *Biol Cell* 66:145-148.
- 45 Garrick RA, Polefka TG, Cua WO, Chinard FP (1986) Water permeability of alveolar macrophages. *Am J Physiol Cell Physiol* 251:C524-C528.
- 46 Garrick RA, Ryan US, Chinard FP (1988) Water permeability of isolated endothelial cells at different temperatures. *Am J Physiol Cell Physiol* 255:C311-C314.
- 47 Folkesson HG, Matthay MA, Frigeri A, Verkman AS (1996) Transepithelial water permeability in microperfused distal airways. Evidence for channel-mediated water transport. *J Clin Invest* 97:664-671.
- 48 Squier CA, Cox P, Wertz PW (1991) Lipid content and water permeability of skin and oral mucosa. *J Invest Dermatol* 96:123-126.
- 49 Whitley V (1998) Influence of unstirred layers on membrane transport in the mammalian kidney. State University of New York at Stony Brook.

- 50 Field MJ, Young JA (1973) Kinetics of Na transport in the rat submaxillary main duct perfused *in vitro*. Pflügers Arch 345:207-220.
- 51 Layton HE, Knepper MA, Chou CL (1996) Permeability criteria for effective function of passive countercurrent multiplier. Am J Physiol Renal Physiol 270:F9-F20.
- 52 Layton HE (1986) A mathematical model of the urine concentrating mechanism (kidney, counter-current system, integral equations); (Doctoral dissertation, Duke University).
- 53 Jen JF, Stephenson JL (1994) Externally driven countercurrent multiplication in a mathematical model of the urinary concentrating mechanism of the renal inner medulla. Bull Math Biol 56:491-514.
- 54 Wexler AS, Kalaba RE, Marsh DJ (1987) Passive, one-dimensional countercurrent models do not simulate hypertonic urine formation. Am J Physiol Renal Physiol 253:F1020-F1030.
- 55 Nawata CM, Pannabecker TL (2018) Mammalian urine concentration: a review of renal medullary architecture and membrane transporters. J Comp Physiol B 188:899-918.